

ARGOPHYLLONE-B, A SESQUITERPENE LACTONE FROM *HELIANTHUS ARGOPHYLLUS*

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Key Word Index—*Helianthus argophyllus*; germacranolide; argophyllone-B; X-ray crystal structure; sesquiterpene lactone.

Abstract—The sesquiterpene lactone, 2-methyl-2-butenic acid dodecahydro-4-(hydroxymethyl)-10a-methyl-8-methylene-3,7-dioxoxineno[5,6]cyclodeca[1,2-*b*]furan-9-yl ester [1aR*-[1aS*,4R*,5aS*,8aR*,9R*(*E*)]], argophyllone-B, was isolated from acetone extracts from the leaves of *Helianthus argophyllus*. Its structure has been determined by single crystal X-ray analysis. Complete ^1H NMR and ^{13}C NMR assignments have been made.

INTRODUCTION

Helianthus species are known to be a rich source of terpenes, including diterpenes and sesquiterpene lactones (for a review see ref. [1]; [2–9]). During our investigation of natural products in sunflower, we isolated a new compound from *H. argophyllus*. Previous work on this species has been reported [2]. This paper reports the isolation and structure of the sesquiterpene lactone, argophyllone-B (1).

RESULTS AND DISCUSSION

Acetone extracts from freeze-dried leaves of *H. argophyllus* yielded a sesquiterpene lactone as indicated in the ^1H NMR spectrum by two one-proton doublets at $\delta 6.37$ ($J = 1.3$ Hz) and 5.84 ($J = 1.3$ Hz) characteristic of the vinyl protons of an α -methylene-lactone (Table 1). The compound contained an angelic acid ester as shown by ^1H NMR peaks at $\delta 6.13$ (1H, q of q , $J = 1.4$ Hz), 1.97 (3H, d of q , $J = 7.3$ and $J = 1.5$ Hz) and 1.84 (3H, pentet, J

Table 1. ^1H NMR spectral data for argophyllone-B (360 Mz, CDCl_3 , CDCl_3 as internal standard)

Proton	δ	Multiplicity (J in Hz)
1	3.39	<i>dd</i> (9.8, 5.6)
2 α	2.35	<i>dd</i> (14.4, 9.8)
2 β	3.29	<i>dd</i> (14.4, 5.6)
4	3.21	<i>m</i>
5 α	1.53	<i>ddd</i> (13.2; 11.4, 3.3)
5 β	2.23	<i>td</i> (13.2, 3.3)
6	4.56	<i>dd</i> (11.4, 3.3)
7	3.06	<i>s</i> (<i>br</i>)
8	5.22	<i>t</i> (≈ 3.5)
9 α	1.45*	<i>dd</i> (15.4, 3.9)
9 β	2.72*	<i>dd</i> (15.4, 3.3)
13 α	5.84	<i>d</i> (1.3)
13 β	6.37	<i>d</i> (1.3)
14	1.40‡	<i>s</i>
15 α	3.87†	<i>dd</i> (9.7, 8.7)
15 β	3.69†	<i>dd</i> (9.7, 4.9)
18	6.13	<i>qq</i> (7.3, 1.4)
19	1.97‡	<i>dq</i> (7.3, 1.5)
20	1.84‡	pentet (1.4)

*†Assignments carrying the same symbol are interchangeable.

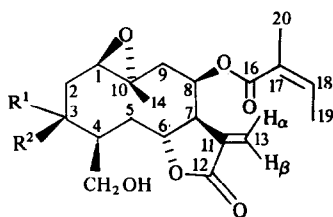
‡Three protons.

Table 2. ^{13}C NMR spectral data for argophyllone-B (22.6 MHz)

Carbon	δ		Multiplicity (in coupled spectrum)
	CDCl_3	C_6D_6	
1	58.7	58.5	<i>d</i>
2	45.8	46.2	<i>t</i>
3	209.4	208.9	<i>s</i>
4	46.3	46.2	<i>d</i>
5	34.8	35.1	<i>t</i>
6	*	75.5†	—
7	47.9	47.5	<i>d</i>
8	*	76.2†	—
9	43.5	43.8	<i>t</i>
10	57.3	57.0	<i>s</i>
11	137.3	138.4	<i>s</i>
12	168.6	168.3	<i>s</i>
13	125.4	124.3	<i>t</i>
14	18.0	17.9	<i>q</i>
15	65.0	65.4	<i>t</i>
16	165.8	165.8	<i>s</i>
17	126.3	*	<i>s</i>
18	141.1	141.2	<i>d</i>
19	15.6	15.8	<i>q</i>
20	20.1	20.4	<i>q</i>

*Signal obscured by solvent peak.

†These assignments are interchangeable.



1 $R^1 = R^2 = O$

2 $R^1 = H, R^2 = OH$

= 1.4 Hz). The ^{13}C NMR spectrum supported this assumption, indicating 20 carbons (Table 2).

Although other structural features were apparent from the 1H NMR spectrum, complete assignments were originally complicated because the molecule did not show a parent peak in the mass spectrum (highest m/z at 360, $[M - H_2O]^+$). However, a single crystal X-ray study, by direct methods, clearly showed argophyllone-B to have structure 1, which has been partially refined to $R = 9.2\%$; H atoms have been located in a difference map. (A completely refined structure with absolute configuration determination will be the subject of a separate paper.) Careful analysis of the 1H NMR spectrum, using decoupling experiments, allowed the assignment of all 25 protons attached to carbons. The $-CH_2-OH$ group appeared as two doublets of doublets at $\delta 3.87$ and 3.69 . These methylene protons were coupled to a methine proton (C-4) at $\delta 3.21$. The methine proton was coupled to another methylene group (C-5) which appeared as an overlapping doublet of doublets at $\delta 1.53$ and a triplet of doublets at $\delta 2.23$. The peaks at $\delta 1.53$ and 2.23 were identified as a pair of methylene protons because of their geminal coupling constant ($J = 13.2$ Hz). These methylene protons were coupled to an oxygen-substituted methine (C-6) proton at $\delta 4.56$.

Other couplings could be observed between an oxygen-substituted methine proton (C-1) at $\delta 3.39$ and the methylene protons (C-2) at $\delta 2.35$ and 3.29 . Oxygen-substituted methine-methylene coupling was also observed between the methine (C-8) peaks at $\delta 5.22$ and the methylene protons (C-9) at $\delta 1.45$ and 2.72 . A broadened singlet (1H) was observed at $\delta 3.06$ (C-7) and a sharp singlet (3H) at $\delta 1.40$ (C-14).

Comparison of structure 1 with that of argophyllin-B (2) [2], isolated from the same species, shows that argophyllone-B has a C-3 keto function rather than a C-3 alcohol. Consistent with this close similarity is the agreement between many of the chemical shifts in both the 1H NMR and ^{13}C NMR spectra for the two compounds.

EXPERIMENTAL

1H NMR and ^{13}C NMR spectra were measured at 360 and 22.5 MHz, respectively, using $CDCl_3$ as internal standard ($\delta 7.26$

for 1H and $\delta 76.9$ for ^{13}C). MS were recorded at 70 eV on a Varian CH-7 spectrometer. ORD spectra were recorded on a Cary Model 60 recording spectropolarimeter and optical rotations were determined on a Perkin-Elmer Model 241. Purifications were performed on Mallinckrodt silica gel (column: CC-4; TLC: 7GF).

Extraction and isolation. Freeze-dried and ground leaves (100 g) of *H. argophyllus* (collected in July 1983, along U.S. Highway 281 at Encino, TX) were extracted with Me_2CO (2 l). The soln was concd and absorbed on silica gel (15 g) and chromatographed over a silica gel (120 g) column. The column was eluted with $C_6H_6-EtOAc-MeOH$ (49:49:1), and 24 ml fractions were collected. Argophyllone-B was concd in fractions 15-35. The crude material was reabsorbed on silica gel (5 g) and chromatographed over a silica gel column (50 g). The column was eluted with $CHCl_3$ (1 l) and 22 ml fractions were collected. Argophyllone-B was concd in fractions 39-92. A small sample was purified by TLC ($C_6H_6-EtOAc-i-PrOH$, 24:24:1). The UV absorbing band was eluted with Et_2O and concd. Argophyllone-B crystallized after standing in the cold. These crystals were used as seeds, providing 236 mg of crude argophyllone-B (mp 140-164/ C_6H_6).

Argophyllone-B (1). Mp 171-179° (C_6H_6), MS m/z (rel. int.): (parent ion not observed), 360 (0.8), 261 (2), 243 (7), 215 (6), 173 (6), 145 (6), 98 (6), 97 (11), 95 (12), 91 (7), 84 (8), 83 (100). $IR_{\nu_{max}}^{CDCl_3}$, cm^{-1} : 3625, 2970, 1770, 1720, 1705, (sh), 1270, 1230, 1150, 1125, 1045. $[\alpha]_D^{22} -158^\circ$ (c 0.06; $EtOH$). ORD (c 1.6×10^{-3} ; $EtOH$) 25° ; $[\Phi]_{400} +1.1^\circ \times 10^3$; $[\Phi]_{332} 0.0^\circ$; $[\Phi]_{320} +1.5^\circ \times 10^3$; $[\Phi]_{317} +1.4^\circ \times 10^3$; $[\Phi]_{313} +1.5^\circ \times 10^3$; $[\Phi]_{307} 0.0^\circ$; $[\Phi]_{230} -4.3^\circ \times 10^4$; $[\Phi]_{220} -1.8^\circ \times 10^4$. Crystal data: (140 K) orthorhombic, $P2_12_12_1$, $Z = 4$, $a = 6.741$ (2), $b = 14.963$ (6), $c = 19.000$ (14) Å; current $R = 9.2\%$, isotropic.

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REFERENCES

- Gershenzon, J., Ohno, N. and Mabry, T. J. (1981) *Rev. Latinoam. Quim.* 12, 53.
- Watanabe, K., Ohno, N., Yoshioka, H., Gershenzon, J. and Mabry, T. J. (1982) *Phytochemistry* 21, 709.
- Ferguson, G., McCrindle, R., Murphy, S. T. and Parvez, M. (1982) *J. Chem. Res.* 200.
- Herz, W., Govindan, S. and Watanabe, K. (1982) *Phytochemistry* 21, 946.
- Spring, O., Albert, K. and Hager, A. (1982) *Phytochemistry* 21, 2551.
- Miyazawa, M. and Kameoka, H. (1983) *Phytochemistry* 22, 1040.
- Mitscher, L. A., Rao, G. S. R., Veysoglu, T., Drake, S. and Haas, T. (1983) *J. Nat. Prod.* 46, 745.
- Beale, M. H., Bearder, J. R., MacMillan, J., Matsuo, A. and Phinney, B. O. (1983) *Phytochemistry* 22, 875.
- Herz, W., Kulanthaivel, P. and Watanabe, K. (1983) *Phytochemistry* 22, 2021.